



Lytton John Musselman

HAUSTORIUM

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HAUSTORIUM BY EMAIL AND THE WEB

We are pleased to acknowledge that Old Dominion University is once again contributing to the printing and mailing of Haustorium but apart from this modest assistance the newsletter still has no significant source of funding and we need to reduce costs as much as possible. The great bulk of our costs are for mailing. Many readers are already helping us by receiving Haustorium by Email. We believe many others could do so but we do not have their Email addresses. If you are one of those, do please let Chris Parker know (Email address on the last page). Bear in mind that having an electronic version of the newsletter enables you to 'search'. If you cannot receive Email, or for any reason wish strongly to go on receiving hard copy, you will continue to receive by airmail.

Thanks to arrangement with the Institute of Arable Crops Research, Long Ashton Research Station, Bristol, UK, Haustorium will continue to be available on the web site: www.lars.bbsrc.ac.uk/cropenv/haust.htm

SEVENTH INTERNATIONAL PARASITIC WEED SYMPOSIUM

The 7th International Symposium will be held in Nantes, France from 5-8 June, 2001. Although the title refers to 'Weed', all aspects of parasitic plants will, as always, be covered, including academic and non-agricultural topics. Those who have not already received a copy of the second circular for this major meeting should contact Patrick Thalouarn, Laboratoire de Cytopathologie Vegetale, University de Nantes, 2, Rue de la Houssinière, BP 92208, F44322 Nantes, Cedex 3, France. Email patrick.thalouarn@svt.univ-nantes.fr

PARASITIC PLANT MANAGEMENT IN SUSTAINABLE AGRICULTURE

The proposal under this title, for a 5-year 25 million ECU programme to be funded from the COST budget of the European Commission is to be considered shortly by the COST Technical Committee in Vienna. If approved, the project will involve a wide range of research and co-ordination activities across a number of European countries.

STRIGA CONTROL BY INTERCROPPING WITH DESMODIUM SPECIES

Resource-poor cereal farmers in Kenya are testing intercropping and trap cropping strategies to control damage by stem boring larvae of moth species such as the indigenous noctuid *Busseola fusca* and the introduced pyralid *Chilo partellus*. They are using two grass species, Sudan grass (*Sorghum sudanensis*) and Napier grass (*Pennisetum purpureum*), that act as "traps" by attracting the pests to lay eggs, and two resistant plants, molasses grass (*Melinis minutiflora*) and the legume silverleaf (*Desmodium uncinatum*), which repel adult stem borers. This "push-pull" strategy is being developed as part of a collaborative programme between IACR-Rothamsted and the International Centre of Insect Physiology and Ecology (ICIPE) at Nairobi and Mbita Point on the banks of Lake Victoria.

In 1997, it was noticed that maize intercropped with silverleaf (*D. uncinatum*) or greenleaf (*D. intortum*) suffered far less *Striga hermonthica* infestation than maize in monoculture. These trials were repeated, comparing *Desmodium* species with plants widely recommended as intercropping solutions to *Striga* problems, for example sunn hemp, *Crotalaria* spp., soya bean and cowpea. With the conventional intercrops, either *Striga* infestation was not significantly different from the maize monoculture, as with soya bean, or the *Striga* rating was only reduced by about 50%, as with sunn hemp and cowpea. However, when maize was intercropped with either of the *Desmodium* species, the *Striga* rating was reduced from 2-3 to 0.1 or less. At the same time, there was a statistically significant increase in maize yield of ca. 2 tonnes/hectare.

Desmodium species are nitrogen-fixing legumes and contribute to the nutrition of the crop. If allowed to grow uncontrolled they may compete with the crop, but this can easily be controlled by regular cutting. The mechanism by which these plants, as intercrops, reduce *Striga* infestation so dramatically is now under investigation, with clear evidence of allelopathic effects. It is now imperative to identify the exact mechanisms by which *Desmodium* species interfere with *Striga* development, to ensure that control measures based on these intercropping strategies are robust and reliable and with a view to exploitation in the longer term by means of plant molecular genetics. It can already be

seen that *D. uncinatum* is producing germination stimulation cues in the rhizosphere, and also that there are compounds interfering with haustorial development and colonisation of the maize host.

The intercropping and trap cropping studies are funded by the Gatsby Charitable Foundation, with new support from the Rockefeller Foundation to fund further mode of action studies on the influence of *Desmodium* species on prevention of *Striga* development. The chemistry involved will be elucidated in the near future by collaborators from ICIPE and Rothamsted: A. Hassanali, A.M. Hooper, T.M. Khamis, Z.R. Khan, J.A. Pickett and L.J. Wadhams.

John A. Pickett, IACR-Rothamsted, Harpenden, Herts AL5 2JQ, UK.

A NEW VIRULENT RACE OF *OROBANCHE CRENATA*

Some vetch varieties are known to resist *O. crenata*. In recent years we have noted a few cases where resistant vetch was attacked by *O. crenata*. One could ask whether this happened as a result of the loss of resistance in the vetch, or as a result of the appearance of a new *Orobanche* race that is capable of attacking the resistant varieties. We have confirmed the botanical identity of the *Orobanche* plants using morphological markers and specific DNA markers that are known to be characteristic of *O. crenata*. Further, the susceptibility of three vetch varieties was checked in the lab against normal *O. crenata* populations, and against the crenate broomrape that was collected on resistant vetch. The results were very clear. The two varieties 'sadot' and 'popany' that are commonly known as *Orobanche* resistant were attacked only by *Orobanche* developing from the seeds collected in the resistant field, not by the 'ordinary' *O. crenata*. This is the first recorded evidence of an *O. crenata* race that overcomes known *Orobanche* resistance.

Daniel M. Joel and Vitaly H. Portnoy, Neve-Ya'ar Research Center, Israel.

SOUTHERN AFRICA STRIGA WORKING GROUP

Striga researchers from Botswana, Malawi, Mozambique, Tanzania and Zimbabwe met in Dar es Salaam in May, 2000, to establish a *Striga* Working Group in Southern Africa. *Striga* infestation is a widespread constraint to cereal productivity in the twelve member states of the Southern Africa Development Community (SADC). The species of economic importance are *S. hermonthica* which occurs in northern Tanzania, *S. asiatica* which is distributed through out the SADC region, and *S. forbesii* which is of local significance in Malawi, Tanzania and Zimbabwe. The meeting, co-ordinated by the SADC/ICRISAT Sorghum and Millet Improvement Program, formulated national action plans for future research and proposed a regional plan involving the working group in a co-ordinated approach to the development, testing and promotion of appropriate *Striga* management options. It was recognised that considerable progress has been made by individual countries to develop *Striga* control components and it was agreed that the emphasis should now be on using available technologies on parasite infested land in integrated crop management which is tested and promoted by farmers themselves. Specific topics which still need attention include filling in gaps in knowledge of *Striga* distribution, and raising awareness further of the importance of the pest at a number of levels of the agricultural community including research managers. Exchange of germplasm and in particular the exposure of farmers to resistant cereal lines needs to be accelerated and attention given to sustainable systems of seed multiplication and distribution. Work is also needed to develop appropriate 'learning tools' which can assist in the dissemination of what tend to be 'knowledge intensive' technologies for *Striga* management, for example improvement of soil fertility or inter-cropping. Such tools should be aimed at farmers and in-service or college training of extension department or NGO agricultural support staff. The working group also hopes to establish a bibliography of previous work on *Striga* in Southern Africa as a resource for researchers in the region. The proceedings of the meeting will be available later in the year.

Charlie R. Riches, Natural Resources Institute, IACR-Long Ashton Research Station, Bristol, BS18 9AF, UK.

STRIGA BIOCONTROL - OBSTACLES OVERCOME?

This is the title of a very encouraging paper presented at the 3rd International Weed Science Congress by Alan K. Watson, Marie Ciotola and Roger R. Maclean. The following is the full abstract (number 371) reproduced with kind permission of the authors, from pp. 176-177 of the Congress Abstracts (see below for a full listing of relevant abstracts, under Proceedings of Meetings.)

'*Fusarium oxysporum* isolate M12-4A is currently being evaluated for the biological control of *Striga hermonthica*. In field trials, chlamydospore powder harvested from small-scale fermentors reduced *S. hermonthica* emergence by 92%. Complete inhibition of *S. hermonthica* emergence occurred when the chlamydospore powder was added to the soil at sowing and when sorghum seeds coated with chlamydospores were sown. Effective biological control of *S. hermonthica* was achieved using a simple fermentation system with sorghum straw as the inoculum using arabic gum as the adhesive. This simple delivery system permits a uniform inoculation of the field as well as the proper positioning of the inoculum in the immediate environment of sorghum roots, where *S. hermonthica* attaches to its host. To facilitate broad usage of *F. oxysporum* M12-4A for the biocontrol of *S. hermonthica*, we are promoting an inoculum production strategy based on a cottage industry model that utilizes a liquid fermentation process and inexpensive locally available substrates including sorghum straw and arabic gum. To assure quality control, primary inoculum is produced centrally and encapsulated in small gelatin capsules as starter cultures. Each capsule contains 0.001 g of inoculum

and a kg box of capsules is sufficient to produce *Fusarium* inoculum for treating up to 8,000 hectares of land infested with *Striga*. In the villages traditional cooking pots filled with water and a small amount of ground sorghum straw are sterilized over a fire, allowed to cool, and the starter culture added. The mixture will ferment for 10-14 days, the product is then air-dried and ground, and can be stored for several months. When planting season arrives, the farmers' cereal seeds are coated with a thin film of arabic gum solution and the dry powdered *Fusarium* inoculum sprinkled onto the seed surface. The biocontrol becomes a 'seed technology'. The farmer plants his seeds and at the same time protects his crop from the ravages of *Striga*. The application rate of *Fusarium* is equivalent to approximately 80 grams per hectare. The village-level manufacture of *Fusarium* will give women in rural communities more economic and social power. The preparation of the dried inoculum both fits into women's traditional sphere of work and provides a new source of income. This year, field testing of the *Fusarium-Striga* biocontrol process is occurring in six Malian villages. We plan to phase this technology in gradually, from village to village and then from country to country, to include all regions afflicted with *Striga*.'

WEBSITES

For information on the 7th International Parasitic Weed Symposium at Nantes, 2001 see:

<http://www.sciences.univ-nantes.fr/scnat/biologie/GPPV.web>

(N.B. notbiologie/scnat...as indicated in the first circular.)

For information on biology and control of parasitic weeds, and the relevant activities of the University of Hohenheim see: <http://www.uni-hohenheim.de/~www380/parasite/start.htm>

For IITA *Striga* Research Methods: a Manual, see: <http://www.cgiar.org/iita> (N.B. www omitted in last issue.)

For news from Canada of progress with biocontrol techniques for *Striga* see: <http://www.mcgill.ca/media/releases/1999/december/weedkiller/>

PROCEEDINGS OF MEETINGS

Breeding for *Striga* Resistance in Cereals. Proceedings of a Workshop held at IITA, Ibadan, Nigeria, from 18-20 August 1999. 2000. Edited by Haussmann, B.I.G., Hess, D.E., Koyama, M.L., Grivet, L., Rattunde, H.F.W. and Geiger, H.H. Available from Margraf Verlag, P.O. Box 105, 97985 Welkersheim, Germany (Email margraf@compuserve.com) at DM 50.- per copy.

Contents:

Gurney A.L. et al. Physiological processes during *striga* infestation in maize and sorghum. pp. 3-17.

Heller, R. and Wegmann, K. Mechanisms of resistance to *Striga hermonthica* (Del.) Benth. in *Sorghum bicolor* (L.) Moench. pp. 19-28.

Ejeta, G. et al. Selection for specific mechanisms of resistance to *striga* in sorghum. pp. 29-39.

Haussmann, B.I.G. et al. Diallel studies on *striga* resistance in sorghum. pp. 41-58.

Omanya, G.O. et al. Evaluation of laboratory, pot, and field measures of *striga* resistance in sorghum. pp. 59-72.

DeVries, J. The inheritance of *striga* reactions in maize. pp. 73-81.

Rattunde H.F.W. et al. Breeding sorghum for *striga* resistance at ICRISAT: progress and perspectives. pp. 85-93.

Gupta, S.C. and Lagoke S.T.O. Transfer of *striga* resistance genes into elite sorghum breeding lines in Nigeria. pp. 95-102.

Kling, J.G. et al. *Striga* resistance breeding in maize. pp. 103-118.

Kabambe, V.H. et al. Screening of teosinte-derived materials for resistance and adaptation to *Striga asiatica* in Malawi, 1998/99 season. pp. 119-125.

Odongo O.M. et al. Screening of teosinte-derived maize lines for resistance to *Striga hermonthica* in Western Kenya. pp. 127-137.

Johnson D.E. et al. The potential for host resistance to *striga* on rice in West Africa. pp. 139-145.

Wilson, J.P. et al. Resistance to *Striga hermonthica* in the primary gene pool of *Pennisetum glaucum*. pp. 147-156.

Haussmann, B.I.G. et al. QTL for *striga* resistance in sorghum populations derived from IS 9830 and N 13. pp.

159-171.

Ejeta, G. Molecular mapping of striga resistance genes in sorghum. p. 173.

Melake-Berhan A. et al. Application of molecular markers for mapping striga resistance gene(s) in maize. pp. 175-185.

Grimanelli, D. et al. Identification of genes for tolerance to striga in maize using transposable elements. p. 187.

Kanampiu, F.K. et al. Utilization of herbicide resistance to combat striga in maize. pp. 189-196.

Bennetzen, J.L. et al. The study and engineering of resistance to the parasitic weed striga in rice, sorghum and maize. pp. 197-205.

Gressel J. and Levy, A. Giving Striga hermonthica the DT's. pp. 207-224.

Koyama, M.L. Molecular markers for the study of pathogen variability: implications for breeding resistance to Striga hermonthica. pp. 227-245.

Koyama, M.L. Genetic variability of Striga hermonthica and effect of resistant sorghum cultivars on population dynamics. pp. 247-260.

Singh, B.B. Breeding cowpea varieties with combined resistance to different strains of Striga gesnerioides. pp. 261-270.

Dashiell, K. et al. Breeding for integrated management of Striga hermonthica. pp. 273-281.

Ado, S.G. et al. Breeding maize for tolerance and resistance to striga at the Institute for Agriculture Research Samaru, Nigeria. pp. 285-290.

Alabi, S.O. et al. Reaction of maize varieties to Striga hermonthica and Striga aspera in the Sudan savanna ecology. pp. 291-298.

Ayiecho, P.O. and Nyabundi, J. Field screening of sorghum cultivars for striga resistance. pp. 299-304.

Belete, K. Status of striga research in Ethiopia. pp. 305-306.

Ebiyau, J. et al. Striga research activities in sorghum at Serere Agricultural and Animal Production research Institute (SAARI), Uganda. pp. 307-311.

Kabambe, V.H. et al. Development of maize genotypes resistant or tolerant to Striga asiatica in Malawi. pp. 313-323

Lagoke, S.T.O. et al., Host plant resistance for striga control in sorghum - activities at IAR, Samaru, Nigeria. pp. 325-334.

Mangombe, N. et al. Breeding sorghum for striga resistance in Zimbabwe. p. 335.

Nour Eldin, I. Screening for striga resistance in sorghum in Sudan. p. 337.

Odhambo G. Striga research activities at KARI, Kenya. p.339.

Ouédraogo, O. et al. Striga research activities in Burkina Faso. p. 341.

Sallah, P.Y.K. et al. Progress in breeding maize for tolerance to Striga hermonthica in Ghana. pp. 343-356.

Future striga research

Application of molecular marker technologies by NARS

List of participants

Group photograph

List of abbreviations

Glossary of genetic terms

(see Haustorium 36, pp. 6-8, for a detailed report of this meeting.)

Striga distribution and management in Tanzania. Proceedings of a stakeholder workshop, Dar es Salaam, September 1999. 2000. Edited by C.R. Riches. Available from Pest Management Dept., Natural Resources Institute, Chatham,

Kent ME4 4TB, UK.

Contents:

Kirway, M.T. Opening address. pp. 1-2.

Mbwaga, A.M. et al. Striga in Tanzania: species distribution and previous work. pp. 3-12.

Mafuru, J.M. The extent of farmer perceptions of Striga in the Lake Zone, Tanzania. pp. 13-16.

Manyerere, A.P. Finger millet production in Serengeti District. pp. 17-18.

Mwambungu, A.H. Status of Striga and Rhamphicarpa in Kyela District. p. 19.

Mpalanga, B. Status of Striga in the Southern Zone. pp. 20-21.

Lamboll, R. The Striga in Dodoma Region: analysis of the problem and research priorities. pp. 22-30.

Press, M.C. et al. Key concepts underpinning Striga control. pp. 31-33.

Mbwaga, A.M. Evaluation of sorghum for Striga resistance. pp. 34-37.

Lamboll, R. Sorghum variety preference - Dodoma. pp. 38-41.

Kapinga, E.B. Research strategies for Striga control in the Lake Province of Tanzania. pp. 42-46.

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Gurney, A.L. et al. Physiological responses of sorghum and maize to infection by Striga. pp. 48-51.

Riches, C.R. Stimulant production by potential trap crops. p. 52.

Press, M.C. et al. Control of Striga on maize and sorghum: nitrogen x crop genotype interactions. pp. 53-55.

Mbwaga, A.M. On-farm evaluation on the use of animal manure to control Striga. pp. 56-57.

Ley, G.L. Opportunities for enhancing soil fertility in smallholder systems in Tanzania. pp. 58-60.

Lamboll, R. Options for Striga control in rice cropping systems in Kyela. pp. 61-64.

Mbwaga, A.M. Screening of local and exotic rice germplasm for Striga asiatica in upland rice. pp. 65-66.

Kayeke, J. On farm evaluation of the use of urea to control Striga asiatica in upland rice. pp. 67-68.

Kaswende, J. Development of Striga control options in maize. pp. 69-74.

Scholes, J.D. et al. Novel sources of resistance to Striga spp. in sorghum and maize. pp. 75-79.

Riches, C.R. Future Striga research priorities for Tanzania. pp. 80-85.

XIVth International Plant Protection Congress - Plant Protection towards the Third Millenium: when Chemistry meets Ecology. Jerusalem, July 1999.

Relevant abstracts include:

Joel, D.M. Long-term approach for parasitic weeds control: manipulation of specific developmental mechanisms of the parasite. (p. 44)

Plakhine, D. et al. Effect of imidazolinone herbicides on early development stages of *Orobanche aegyptiaca*. (p. 126)

Kleifeld, Y. Management and control of *Orobanche* and *Cuscuta*. (p. 45)

Antonova, T.S. Review of common traits in sunflower resistance to different pathogens. (p. 45)

Eizenberg, H. et al. Resistance mechanisms of sunflower (*Helianthus annuus*) to *Orobanche cumana*. (p. 77)

Goldwasser, Y. et al. Anatomical studies of *Vicia atropurpurea* resistance to *Orobanche aegyptiaca*. (p. 81)

Ransom, J. Cultural, chemical and biological control of Striga in sustainable agriculture. (p. 45)

Oswald, A. et al. On-farm research and training of farmers' groups on Striga control using a participative approach.

Oswald, A. et al. Crop rotation to reduce Striga and increase overall productivity in maize-based cropping systems. (p. 74)

Odhambo, G.D. and Ransom, J. Effect of organic and inorganic sources of nitrogen on control of Striga hermonthica and on soil fertility for higher maize productivity in Western Kenya. (p. 73)

Abayo, G.O. et al. Effect of short-term improved fallow on Striga infestation in maize. (p. 103)

Kanampui, F.K. et al. Herbicide seed dressings of corn bearing ALS target-site resistance with ALS-inhibiting herbicides for witchweed control. (p. 122)

Bedi, J.S. et al. Efficacy of a Fusarium oxysporum formulation for the control of Orobanche cumana. (p. 70)

Weinberg, Ts. et al. Response of Cuscuta campestris to herbicide inhibitors of carotenoid biosynthesis. (p. 117)

Nof, E. et al. Biological control of field dodder by a pathogenic fungus. (p. 109)

N.B. Abstracts of these papers are usefully reprinted in *Phytoparasitica* (2000) 28: 171-177.

Third International Weed Science Congress - IWSC. Foz do Iguassu - June 6 to 11, 2000. Abstracts.

Edited by Anne Légère. 301 pp.

Relevant abstracts include:

(abstract number in brackets)

Goldwasser et al. Control of Orobanche spp. in potato. (278) pp 131-132.

Kanampui et al. Herbicide seed dressing of corn as an appropriate treatment for Striga control while allowing intercropping. (282). pp. 133-134.

Canevari, M. and Colbert, D. Post emergence control of Cuscuta pentagona in alfalfa hay. (288). pp. 136-137.

Kroschel, J. and Klein, O. Natural impact and potential for biological control of two selected antagonists of the parasitic weeds Orobanche spp. and Cuscuta spp. (366). p. 174.

Watson, A.K. et al. Striga biocontrol - obstacles overcome. (371). pp. 176-177. (NB. see full abstract above.)

Elzein, A. et al. Effect of storage temperature, granule size, and inoculum type on the viability of Fusarium oxysporum, a pathogen of Striga hermonthica, encapsulated in wheat-kaolin ("Pesta") granules. (377). pp. 179-180.

Braghouthi et al. Effect of bacteria on broomrape seed germination. (385). pp. 183-184.

Riches, C.R. Improved weed management for resource poor farmers: constraints and opportunities. (392). p. 187.

Kroschel, J. Striga: a joint challenge to science, extension and farmers. (395). pp. 188-189.

Eplee, R.E. and Norris, R. Eradication of Striga asiatica from the United States. (442). p. 212.

Labrada, R. An overview of parasitic weed control. (538). p. 260.

Gbèhounou, G. - Striga control by restoring soil fertility. (539). pp. 260-261.

Odhambo, G.D. Progress on Striga hermonthica control in East Africa. (540) p. 261.

Joel, D.M. Prospects for Orobanche control in the 21st century. (541). pp. 261-262.

Oswald, M. et al. Interaction of plant growth promoting rhizobacteria (PGPR) with maize and Striga hermonthica (Del.) Benth. seeds. (543) p. 262.

Oswald, A. et al. Linking research, extension and farmers: Striga control strategies for western Kenya. (544.) p. 263.

Saghir, A.R. New possibilities for Cuscuta management in some vegetable crops. (545). p. 263.

Babiker, A.G.T. et al. Conditioning, CO₂ and GR24 influence ethylene biosynthesis and germination of Striga hermonthica. (546). p. 264.

Ndung'u, D.K. et al. Effect of fodder legumes on stimulation, attachment and emergence of Striga hermonthica on maize. (547). p. 264.

LITERATURE

- Abanvou, L. and Doku, E.V. 1998. Heritability of traits associated with striga (*Striga hermonthica* (Del.) Benth.) resistance in an open-pollinated maize population. *African Crop Science Journal* 6: 129-135.
- Adler, L.S. 2000. Alkaloid uptake increases fitness in a hemi-parasitic plant via reduced herbivory and increased pollination. *The American Naturalist* 156: 92-99. (Castilleja indivisa grown on near-isogenic lines of *Lupinus albus* with low and high levels of alkaloid. Presence of lupanine in *C. indivisa* growing on high-alkaloid *L. albus* reduced damage from larvae of *Junonia coenia* and other moth larvae and increased seed set by 50% due to preference of the pollinating humming bird for undamaged plants.)
- Aigbokhan, E.I., Berner, D.K., Musselman, L.J. and Mignouna, H.D. 2000. Evaluation of variability in *Striga aspera*, *Striga hermonthica* and their hybrids using morphological characters and random amplified polymorphic DNA markers. *Weed Research* 40: 375-386. (Results confirm that the two species are genetically distinct. Hybrids are morphologically intermediate but closer to the maternal parents genetically. Some evidence for existence of naturally occurring hybrids in Nigeria.)
- Albrecht, H., Yoder, J.I. and Phillips, D. 1999. Flavonoids promote haustoria formation in the root parasite *Trypophyllum versicolor*. *Plant Physiology* 119: 585-591.
- Amsellem, Z., Zidack, N.K., Quimby, P.C. and Gressel, J. 1999. Longterm dry preservation of viable mycelia of two mycoherbicidal organisms. *Crop Protection* 18: 643-649. (Studies on *Fusarium oxysporum* and *F. arthrosporioides*.)
- Arditti, J. and Ghani, A.K.A. 2000. Tansley Review No. 110. Numerical and physical properties of orchid seeds and their biological implications. *New Phytologist* 145: 367-421. (No mention of parasitic plants but the beautiful illustrations show remarkable parallels with *Orobanche* and *Striga* in their ornamentation and especially with *Alectra* in their tubular structure. See also refs to McKendrick et al. in the same issue.)
- Babiker, A.G.T., Ma, Y., Sugimoto, Y. and Inanaga, S. 2000. Conditioning period, CO₂ and GR24 influence ethylene biosynthesis and germination of *Striga hermonthica*. *Physiologia Plantarum* 109: 75-80. (Results are consistent with a model in which conditioning removes a restriction on the ethylene biosynthetic pathway in *S. hermonthica* seeds: GR24 modulates the key enzymes in ethylene biosynthesis: germination results from joint action of GR24 and the ethylene it induces.)
- Barandiarán, X., Moral, R., Gil, J. and Moreno, M.T. 1999. (The effect of the reduced form of glutathione (GSH) on the germination and growth of *Orobanche crenata*.) (in Spanish) In: SEMh Congreso Sociedad Española de Malherbología, Actas, Logroño, Spain 1999: 145-149.
- Baumgartner, J.R., Al-Khatib, K. and Currie, R.S. 1999. Cross-resistance of imazethapyr-resistant common sunflower (*Helianthus annuus*) to selected imidazolinone, sulphonylurea and triazolopyrimidine herbicides. *Weed Technology* 13: 489-493. (Reporting that the naturally occurring imazethapyr resistant sunflower shows high resistance also to imazamox. cf refs to Alonso et al. in *Haustorium* 35 and Al-Khatib et al. in *Haustorium* 34.)
- Benharrat, H., Delavault, P., Theodet, C., Figureau, C. and Thalouarn, P. 2000. rbcL plastid pseudogene as a tool for *Orobanche* (subsection *Minores*) identification. *Plant Biology* 2(1): 34-39. (Studies of the pseudogene and associated morphological characters lead to a new key for separation of *O. hederaceae*, *O. minor*, *O. amethystea* and *O. loricata*.)
- Bhan, V.M. and Sushi Kumar. 1998. Weed science research in India. *Indian Journal of Agricultural Science* 68: 567-582. (Briefly reviews the limited work in India on management of parasitic weeds by solarisation, cultural, chemical and biological control.)
- Box, J.D. 2000. Mistletoe *Viscum album* L. (Loranthaceae) on oaks in Britain. *Watsonia* 23: 237-256. (Detailed history of *V. album* records on *Quercus* spp. in Britain. Currently just 11 examples are known, mainly on *Q. robur*.)
- Briggs, J. 1999. Kissing goodbye to mistletoe? The results of a national survey aimed at discovering whether mistletoe in Britain is in decline. A joint report from Plantlife - The Wild-Plant Conservation Charity and the Botanical Society of the British Isles. Plantlife, London, 20 pp. (A summary will be included in the next issue.)
- Bringmann, G., Schlauer, J., Rückert, M., Wiesen, B., Ehrenfeld, K., Proksch, P. and Czygan, F.C. 1999. Host-derived acetogenins involved in the incompatible parasitic relationship between *Cuscuta reflexa* (Convolvulaceae) and *Ancistrocladus heyneanus* (Ancistrocladaceae). *Plant Biology* 1: 581-584. (The naphthoquinone plumbagin demonstrated to be one of the factors involved in dieback of *C. reflexa* after attachment to *A. heyneanus*.)
- Calladine, A. and Pate, J.S. 2000. Haustorial structure and functioning of the root hemiparasitic tree *Nuytsia floribunda* (Labill.) R.Br. and water relationships with its hosts. *Annals of Botany* 85: 723-731. (Describing in new detail the structure and function of the 'cutting device' of *L. floribunda* and demonstrating uptake of water from the xylem of the host *Acacia acuminata*.)

Calladine, A., Pate, J.S. and Dixon, K.W. 2000. Haustorial development and growth benefit to seedlings of the root hemiparasitic tree *Nuytsia floribunda* (Lobill.) R.Br. in association with various hosts. *Annals of Botany* 85: 733-740. (Over a 12 month period *N. floribunda* growth was roughly proportional to the number and weight of haustoria developing on roots of a range of 23 woody host species. Some evidence for uptake of N from the nodulated hosts *Acacia cyclops* and *C. acuminata*.)

Carsky, R.J., Berner, D.K., Oyewole, B.D., Dashiell, K. and Schulz, S. 2000. Reduction of *Striga hermonthica* parasitism on maize using soybean rotation. *International Journal of Pest Management* 46: 115-120. (In 2 out of 3 field trials, soybean grown in year 1 resulted in 50-70% lower *S. hermonthica* numbers in maize in year 2 compared with sorghum in year 1. Some evidence for greater effect of soybean at higher densities and with added P to increase root development but as *Striga* was allowed to seed in sorghum, exact benefit difficult to judge.)

Caruso, C.M. and Alfaro, M. 2000. Interspecific pollen transfer as a mechanism of competition: effect of *Castilleja linariaefolia* pollen on seed set of *Ipomopsis aggregata*. *Canadian Journal of Botany* 78: 600-606.

Chatterjee, U. and Sanwal, G.G. 1999. Purification and properties of a protein from *Lantana camara* activating *Cuscuta reflexa* cellulase. *Phytochemistry* 52: 361-366.

Chou ChengJen, Ko HauChieh and Len LieChwen. 1999. Flavonoid glycosides from *Viscum alniformosanae*. *Journal of Natural Products* 62: 1421-1422. (Two new flavonoid glycosides identified from *V. alniformosanae* in Taiwan.)

Ciotola, M., Ditommaso, A. and Watson, A.K. 2000. Chlamyospore production, inoculation methods and pathogenicity of *Fusarium oxysporum* M12-4A, a biocontrol for *Striga hermonthica*. *Biocontrol Science and Technology* 10: 129-145. (Reporting studies on production, longevity and germination of *F. oxysporum* isolate from Mali, as influenced by nutrients, sorghum root exudate, etc. Successful reduction of *Striga* by use of 0.5-1 g chlamyospore powder per hill.)

Conaghan, J. 1998. *Orobanche hederæ* Duby. in East Donegal (H34). *Irish Naturalists' Journal* 269(3/4): 136.

Cullings, K. 2000. Reassessment of phylogenetic relationships of some members of the Monotropoideae based on partial 28S ribosomal RNA gene sequencing. *Canadian Journal of Botany* 78: 1-2. (Reporting results at variance with the author's previous conclusions on the subject.)

Debabrata Das, Ghosh R.B. Avik Dutta and Maji, U.K. 1999. A census on the hosts of *Cuscuta reflexa* Roxb. in the district of lateritic Bankura, West Bengal. *Environment and Ecology* 17: 763-764. (Fifteen host species listed.)

Debabrata Das, Ruma Hazra, Avik Dutta and Maji, U.K. 1999. Systematic enumeration and taxonomic survey of host-plants of *Cuscuta reflexa* Roxb. in Purulia district, West Bengal. *Environment and Ecology* 17: 479-480. (Of 32 hosts listed from 22 families, most were trees or shrubs and only one a monocot.)

Deeks, S.J., Shamoun, S.F. and Punja, Z.K. 1999. Tissue culture of parasitic flowering plants: methods and applications in agriculture and forestry. *In Vitro Cellular Development and Biology - Plant* 35: 369-381. (A comprehensive review of the literature covering tissue culture of parasitic plants from 23 genera in 7 families and discussing the potential value of such studies.)

Deliorman, D., Calis, I., Ergun, F. and Tamer, U. 1999. The comparative studies on phenylpropanoid glycosides of *Viscum album* subspecies by high performance liquid chromatography. *Journal of Liquid Chromatography and Related Technologies* 22: 3101-3114. (Comparing content of syringin, coniferin and kalopanaxin D in the 3 subspecies *album*, *abietis* and *austriacum* of *Viscum album* in Turkey.)

de Luque, A.P., Galindo, J.C.G., Macias, F.A. and Jorin, N. 2000. Sunflower sesquiterpene lactone models induce *Orobanche cumana* seed germination. *Phytochemistry* 58: 45-50. (Parthenolide and a related lactone stimulate germination of *O. cumana* at 1 mg/l but did not stimulate *O. crenata*, *O. ramosa* or *O. aegyptiaca*.)

Díaz-Sánchez, J., López-Granados, F. and García-Torres, L. 1999. (Absorption and translocation of propyzamide applied to the crop seed, and of imazapyr and glyphosate applied post-emergence to sunflower, as affected by parasitism of nodding broomrape (*Orobanche cumana*.) (in Spanish) In: SEMh Congreso Sociedad Española de Malherbología, Actas, Logroño, Spain 1999: 329-333.

Elzein, A.E.M., Kroschel, J., Assefa Admasu and Masresha Fetene. 1999. Preliminary evaluation of *Phytomyza orobanchia* (Diptera: Agromyzidae) as a controller of *Orobanche* spp. in Ethiopia. *Sinet, an Ethiopian Journal of Science* 22(2): 271-282. (At one location, Matima, *P. orobanchia* found to destroy 81 and 72% of capsules of *O. ramosa* and *O. cernua* respectively, on tomato. It was not found at 2 other sites.)

Erenst, M. and Scheffler, A. 1999. Photohaemolytic activity of chlorophyll degradation products in a mistletoe extract. *Planta Medica* 65: 627-631. (Involving *Viscum album*.)

Foley, M.J.Y. 2000 A morphological comparison between some British *Orobanche* species (*Orobanchaceae*) and their closely-related non-British counterparts from continental Europe: *Orobanche reticulata* Wallr. s.l. *Watsonia* 23:

257-267. (Concluding that *O. reticulata* s.l. from lowland Britain is distinct from, but close to that from continental European mountain areas, and suggesting that the British form be referred to as ssp. *procera* (Koch) Dostál.)

García, M.A. 1999. *Cuscuta* subgenus *Cuscuta* (Convolvulaceae) in Ethiopia, with the description of a new species. *Annales Botanici Fennici* 36: 165-170. (A study of the *Cuscuta* spp. in Ethiopia concluded that *C. approximata* and *C. pedicellata* do not occur, but a new endemic species *C. castroviejoi* is described.)

Gebremedhin, W., Goudriaan, J. and Naber, H. 2000. Morphological, phenological and water-use dynamics of sorghum varieties (*Sorghum bicolor*) under *Striga hermonthica* infestation. *Crop Protection* 19: 61-68. ((Detailed comparison of susceptible var. IS9302 and resistant var. SRN39 in pots.)

Goldwasser, Y., Plakhine, D., Kleifeld, Y., Zamski, E. and Rubin, B. 2000. The differential susceptibility of vetch (*Vicia* spp.) to *Orobancha aegyptiaca*: anatomical studies. *Annals of Botany* 85: 257-262. (Germination of *O. aegyptiaca* and penetration into the cortex occurs in both susceptible *V. sativa* and resistant *V. atropurpurea* but in the latter there is no penetration of the endodermis, associated with production of an unidentified secretion.)

Goldwasser, Y., Plakhine, D. and Yoder, J.I. 2000. *Arabidopsis thaliana* susceptibility to *Orobancha* spp. *Weed Science* 48: 342-346. (*A. thaliana* shown to stimulate germination and support development of *O. aegyptiaca*, *O. ramosa* and *O. minor* but not *O. crenata* and *O. cumana*.)

González-Andújar, J.L., Martínez-Cob, A., López-Granados, F. and García-Torres, L. 1999. (Spatial distribution of crenate broomrape (*Orobancha crenata*) in faba bean: geostatistical analysis.) (in Spanish) SEMh Congreso 1999: Sociedad Española de Malherbología, Actas, Lagroño, Spain, 1999, pp. 139-143. (Results of spherical modelling revealed an aggregated distribution of *O. crenata* population with a patch size of 10 m.)

Gowda, B.S., Riopel, J.L. and Timko, M.P. 1999. NRSA-1: a resistance gene homolog expressed in roots of non-host plants following parasitism by *Striga asiatica* (witchweed). *Plant Journal* 20: 217-230. (Growth of *S. asiatica* was blocked in cortex of non-host *Tagetes minuta* with development of browning and necrosis: this was associated with expression of the nuclear gene NRSA-1 which showed relationship to genes for disease resistance in other species.)

Gworgwor, N.A., Anaso, A.B.J., and Turaki, Z.G. 1998. Integrated cultural practices for *Striga* control in millet-based cropping systems. In: Emechebe, A.M., Ikwelle, M.C., Ajayi, O., Aminu-Kano, M. and Anaso, A.B.J. (Eds.) *Pearl Millet in Nigerian Agriculture: Production, Processing and Research Priorities*, pp. 87-97. (Discussing a range of cultural practices.)

Haidar, M.A. and Sidahmed, M.M. 2000. Soil solarization and chicken manure for the control of *Orobancha crenata* and other weeds in Lebanon. *Crop Protection* 19: 169-173. (Effect of chicken manure apparently greater than that of solarization on *O. crenata* on faba bean in pots, but methodology not clear, and fate of faba bean not reported.)

Hausmann, B.I.G., Hess, D.E., Koyama, M.L., Grivet, L., Rattunde, H.F.W. and Geiger, H.H. 2000. Breeding for *Striga* Resistance in Cereals. *Proceedings of a Workshop held at IITA, Ibadan, Nigeria, from 18-20 August 1999*. ICRISAT/Margraf Verlag, Germany, 376 pp. (See above for availability and a list of contents, and *Haustorium* 36 for a detailed report of this meeting.)

Hausmann, B.I.G., Hess, D.E., Welz, H-G. and Geiger, H.H. 2000. Improved methodologies for breeding *striga*-resistant sorghums. *Field Crops Research* 66: 195-211. (An extended review article.)

Hess, D.E. and Fernández-Rivera, S. 2000. A note on the chemical composition, intake and digestion of *Striga hermonthica* herbage by sheep. *Weed Research* 40: 351-358. (*S. hermonthica* had relatively high levels of N and P but possible problems from high ash and seed transmission need to be addressed before promoting utilization of hand-pulled plants by sheep.)

Hollier, J. and Briggs, J. 1999. The specialist Hemiptera associated with mistletoe. *British Journal of Entomology and Natural History* 12: 237-239. (Species collected included the psyllid *Psylla visci*, the mirid *Orthops visciicola* and the tortricid *Celypha woodiana*; also the cimicid *Anthocoris visci*, a predator feeding only on *P. visci*.)

Huhta, A-P., Tuomi, J. and Rautio, P. 2000. Cost of apical dominance in two monocarpic herbs, *Erysimum strictum* and *Rhinanthus minor*. *Canadian Journal of Botany* 78: 591-599. (Apical damage had negligible effect on vegetative biomass or fecundity of *R. minor*.)

IDRC. 2000. Biological warfare. IDRC Annual Report 1998-1999. pp. 36-39. (Giving a very encouraging summary of progress in the use of *Fusarium oxysporum* for control of *Striga hermonthica* in Mali, including the production of the inoculum at the village level.)

Ihl, B. and Wiese, K. 2000. (Studies on *Cuscuta reflexa* Roxb.: VIII. Mechanical induction of haustoria formation in non-twining stems of the parasite.) (in German) *Flora (Jena)* 195(1): 1-8. (Haustoria could be induced on the sub-apical zone of the stem. The possible involvement of an interaction between IAA and cytokinin is discussed.)

IWSS. 2000. Abstracts, Third International Weed Science Congress - IWSC, Foz do Iguassu, Brazil, 2000. International Weed Science Society. 301 pp. (See above for listing of relevant abstracts.)

Janssen, T. and Wulf, A. 1999. (On the significance of mistletoes for forest protection.) (in German) Mitteilungen aus der Biologischen Bundesanstalt für Land- und Forstwirtschaft, Heft 369, 142pp. (*Loranthus europaeus* is only of importance in parts of Austria; forms of *Viscum album* are more widespread, the most important being *V. album* var. *abietis*. *Arceuthobium* spp. are of no significance in Europe at present but the extreme damage they cause in N. America fully justifies the current quarantine legislation in Europe.)

Joel, D.M., Aviv, D., Surov, T., Portnoy, T., Goldman-Guez, T. and Gressel, J. 1999. Transformation of crops to herbicide-resistance and their use against parasitic weeds. In: Altman, A. et al., (eds.) Plant Biotechnology and in vitro Biology in the 21st Century. Kluwer Academic Publishers, The Netherlands, pp. 499-502. (Reporting promising control of *Orobancha aegyptiaca* with asulam applied post-emergence to asulam-resistant potato; also with glyphosate applied as a seed dressing to glyphosate-resistant rapeseed.)

Joel, D.M., Herschenhorn, J., Goldman-Guez, T., Cohen, E., Lovan, Y. and Portnoy, V.H. 2000. Grafted host plants for broomrape (*Orobancha* spp.) control. Abstracts, Weed Science Society of America, 40: 71-72. (Susceptible sunflower and pepper, grafted onto resistant root-stocks were fully protected.)

Joel, D.M., Plakhine, D., Creanje, P., Dupuis, J.M., Kamodo, J. and Gressel, J. 2000. Broomrape (*Orobancha aegyptiaca* Pers.) control using asulam-resistant seeds coated with asulam. Abstracts, Weed Science Society of America, 40: 57. (Asulam-resistant tobacco grew normally in *Orobancha*-infested soil, while unprotected plants died.)

Juan, R., Pastor, J. and Fernández, I. 2000. SEM and light microscope observations on fruit and seeds in Scrophulariaceae from Southwest Spain and their systematic significance. *Annals of Botany* 86: 323-338. (Includes key for identification to genus level via fruit/seed characteristics. Parasitic genera covered include *Bellardia* (SEM of seed of *B. trixago*), *Parentucellia*, *Bartsia*, *Odontites* and *Pedicularis*.)

Kanampiu, F.K., Friesen, D.K., Ransom, J.K. and Gressdel, J. 2000. Intercropping is not precluded when ALS herbicide-coated corn seed is used for controlling *Striga*. Abstracts, Weed Science Society of America, 40: 7-8. (No effect on crops planted 15 cm away from maize seed treated with imazapyr or pyrithiobac.)

Karim, S.M.R., Mamun, A.A. and Islam, N. 1999. Agroecology of major crops and their weeds in Bangladesh. *Pakistan Journal of Scientific and Industrial Research* 42: 295-300. (*Striga densiflora* and *Orobancha indica*, 'newly introduced to the country', are severely damaging sugar cane and mustard respectively.)

Karnakowski, W. 1999. (Notifiable weeds and parasitic plant material imported into Poland during 1996-99.) (in Polish) *Ochrona Roślin* 44(12): 15, 33. (Summarising interception of notifiable weeds including *Cuscuta*, *Orobancha* spp.)

Kasembe, E., Chivinge, O.A., Mariga, I.K. and Mabasa, S. 1999. The effect of time of ridging on witchweed (*Striga asiatica* (L.) Kuntze) emergence, density and maize grain yield in the small-holder farming sector of Zimbabwe. *Journal of Plant Protection in the Tropics* 12: 15-26.

Kebreab, E. and Murdoch, A.J. 1999. Predicting *Orobancha* seed longevity for better weed management in legumes. *Grain Legumes* No. 23 - 1st quarter 1999: 8-9. (Longevity of seeds greatly reduced at higher temperatures and/or higher moisture.)

Kebreab, E. and Murdoch, A.J. 2000. The effect of water stress on the temperature range for germination of *Orobancha aegyptiaca* seeds. *Seed Science Research* 10: 127-133. (Optimum germination temperature was 17-26°C at high water potential, somewhat lower, 17-20°C with a decrease in water potential.)

Kelly, C.K. and Horning, K. 1999. Acquisition order and resource value in *Cuscuta attenuata*. *Proc. National Academy of Sciences of the United States of America* 96: 13219-13222. (*C. attenuata* shown to grow more vigorously when parasitising more than one host simultaneously.)

Kim, S.K., Akintunde, A.Y. and Walker, P. 1999. Responses of maize inbreds during development of *Striga hermonthica* infestation. *Maydica* 44: 333-339. (Pot experiments with massive inoculation of *S. hermonthica* seeds suggested that 'tolerance' was correlated with tendency to increased root weight in presence of *Striga*, e.g. in Corn Belt inbred B73 and in inbred Per, based on the perennial *Zea diploperennis*.)

Kranz, B. 1999. (Importance of organic manure for the germination and development of the parasitic flowering plant *Striga hermonthica* (Del.) Benth.) (in German) *Agroecology* No. 2: 130 pp. (A thesis exploring the importance of organic matter in the lower occurrence of *S. hermonthica* in fertile 'compound' fields compared with less fertile 'bush' fields, and concluding that N is more important than OM in reducing germination of *Striga* but OM may have other less direct beneficial effects.)

Kuehn, J.J. 1999. (Long term favourable course of a case of centroblastic-centrocytic non-Hodgkin lymphoma under administration of an extract of mistletoe (*Viscum album*)). (in German) *Deutsche Medizinische Wochenschrift* 124:

1414-1418. (Treatment with the *V. album* extract 'Iscador' over a 12 year period held extensive lymphomas in check. Interruption of treatment led to deterioration.)

Ladley, J. J., Kelly, D. and Robertson, A. W. 1997. Explosive flowering, nectar production, breeding systems and pollinators of New Zealand mistletoes (Loranthaceae). *New Zealand Journal of Botany* 35: 345-360. (The pollination syndromes of 5 species of mistletoe; *Alepis flavida*, *Ileostylus micranthus*, *Peraxilla colensoi*, *P. tetrapetala* and *Tupeia antarctica*, are described.)

Ladley, J. J., Kelly, D. and Norton, D. A. 1997. A guide to hand-planting New Zealand mistletoes (Loranthaceae). *New Zealand Botanical Society Newsletter*, 16-19. (General guide to hand-planting mistletoe seeds.)

Lammi, A., Siikämäki, P. and Salonen, V. 1999. The role of local adaptation in the relationship between an endangered root hemiparasite *Euphrasia rostkoviana* and its host, *Agrostis capillaris*. *Ecography* 22: 145-152. (Little evidence for better performance of *E. rostkoviana* when parasitising local populations of the host *A. capillaris* rather than populations from elsewhere.)

Löffler, C., Czygan, F.C. and Proksch, P. 1999. Role of indole-3-acetic acid in the interaction of the phanerogamic parasite *Cuscuta* and host plants. *Plant Biology* 1: 613-617. (*C. reflexa* on tomato causes elongation of cells in host tissue, apparently due to IAA from the parasite, associated with elongation of epithelial cells in parasite haustorial tissue.)

Lu, Y.H., Melero-Vara, J.M., García-Tejada, J.A. and Blanchard, P. 2000. Development of SCAR markers linked to the gene *Or5* conferring resistance to broomrape (*Orobanche cumana* Wallr.) in sunflower. *Theoretical and Applied Genetics* 100: 625-632. (A contribution to more efficient use of resistance genes in sunflower.)

McKendrick, S.L., Leake, J.R., Taylor, D.L. and Read, D.J. 2000. Symbiotic germination and development of myco-heterotrophic plants in nature: ontogeny of *Corallorhiza trifida* and characterization of the mycorrhizal fungi. *New Phytologist* 145: 523-537.

McKendrick, S.L., Leake, J.R. and Read, D.J. 2000. Symbiotic germination and development of myco-heterotrophic plants in nature: transfer of carbon from ectomycorrhizal *Salix repens* and *Betula pendula* to the orchid *Corallorhiza trifida* through shared hyphal connections. *New Phytologist* 145: 539-548. (Two excellent papers challenging the use of the term 'saprophytic' for the orchid *C. trifida* and confirming the importance of transfer of carbon from the indirect hosts *S. repens* and *B. pendula* via ectomycorrhizal fungi. Should we be treating them as parasitic plants?)

Marinescu, A. and Pacureanu-Joita, M. 1998. (Sunflower wild species - sources for resistance to the parasite *Orobanche cumana* Willd.) (in Romanian) *Probleme de Genetica Teoretica si Aplicata* 30(1/2): 67-72.

Marler, M., Pedersen, D., Mitchell-Olds, T. and Calaway, R.M. 1999. A polymerase chain reaction method for detecting dwarf mistletoe infection in douglas fir and western larch. *Canadian Journal of Forest Research* 29: 1317-1321. (A PCR technique described for detecting *Arceuthobium douglasii* and *A. laricis* in the tissues of hosts *Pseudotsuga menziesii* and *Larix occidentalis*.)

Marshall, K. and Filip, G.M. 1999. The relationship of Douglas-fir dwarf mistletoe (*Arceuthobium douglasii*) to stand conditions and plant associations in the southern Cascade Mountains, Oregon. *Northwest Science* 73: 301-311.

Mathiasen, R., Beatty, J. and Melgar, J. 2000. First report of *Arceuthobium hondurensis* on *Pinus tecunumanii*. *Plant Disease* 84: 372. (In Honduras.)

Mathiasen, R., Melgar, J., Beatty, J. and Parks, C. 2000. First report of *Psittacanthus angustifolius* on *Pinus oocarpus* and *Pinus maximinoi*. *Plant Disease* 84: 203. (In Honduras.) Mathiasen, R., Parks, C., Beatty, J. and Sesnie, S. 2000. First report of *Psittacanthus angustifolius* on pines in Mexico and Guatemala. *Plant Disease* 84: 808. (*P. angustifolius* recorded on *Pinus maximinoi*, *P. oocarpa* and possibly *P. tecunumanii*, but no damage observed.)

Mathiasen, R., Sesnie, S., Calderon, J. and Soto, A. 1999. First report of golden dwarf mistletoe on *Pinus maximinoi*. *Plant Disease* 83: 878. (*Arceuthobium aureum* ssp. *aureum* apparently causing witches brooms on *P. maximinoi* in Guatemala.)

Mauromicale, G., Restuccia, G. and Marchese, M. 2000. Germination response and viability of *Orobanche crenata* Forsk. seeds subjected to temperature treatment. *Australian Journal of Agricultural Research* 51: 579-585. (Freshly imbibed seeds exposed to high temperatures - over 40°C - for 12 or 24 hours suffered large reductions in subsequent germination: tetrazolium tests suggested they were mainly dormant up to 65°C but dead above 70°C.)

Mbwaga, A.M., Kaswende, J. and Shayo, E. 2000. A Reference Manual on *Striga* Distribution and Control in Tanzania. SIDA/FAO - FARMESA Programme, P.O. Box Ilonga, Kilosa, Tanzania. 26 pp. (A very sound, well-illustrated booklet, summarising information on biology and control in clear terms, suitable for other researchers, extension personnel and the more literate farmers.)

- Medel, R. 2000. Assessment of parasite mediated selection in a host-parasite system in plants. *Ecology* 81: 1554-1564. (Spine length shown to be important in susceptibility of individuals of cacti, *Echinopsis chilensis* and *Eulychnia acida* to the mistletoe *Tristerix aphyllus*. The study suggests a possible role for mistletoe attack in selection for spine length in *E. chilensis* but not in *E. acida*.)
- Michi, L. Bouillant, M-L., Rohr, R., Sallé, G. and Bally, R. Physiological and cytological studies on the inhibition of *Striga* seed germination by the plant growth-promoting bacterium *Azospirillum brasilense*. *European Journal of Plant Pathology* 106: 347-351.
- Morozov, I.V., Foy, C.L. and Westwood, J.H. 2000. Small broomrape (*Orobanche minor*) and Egyptian broomrape (*Orobanche aegyptiaca*) parasitization of red clover (*Trifolium pratense*). *Weed Technology* 14: 312-320. (Inoculation of *T. pratense* roots with rhizobacteria increased germination and attachment of *O. minor* but not of *O. aegyptiaca*.)
- Müller-Schärer, H., Scheepens, P.C. and Greaves, M.P. 2000. Biological control of weeds in European crops: recent achievements and future work. *Weed Research* 40: 83-98. (Reviewing the activities of a EU-funded working group on biocontrol of *Orobanche* spp., mainly involving fungi, bacteria and fungal toxins.)
- Musselman, L.J. and Vorster, P. 2000. Finding furtive flowers. *Plant Talk* 21: 38-39. (Describing members of *Hydnoraceae* - *Hydnora* spp. in Africa and *Psospanche* spp. in tropical America - with especially interesting observations on *H. triceps* seen in Namaqualand for only the second time this century, on its host *Euphorbia dregeana*.)
- Nair, K.K.N., Pandalai, R.C., Bhat, K.V., Mathew, G. and Ali, M.I.M. 1999. Botany, wood characteristics and silvicultural techniques of the indigenous timber species, *Grewia tiliaefolia* Vahl. *Annals of Forestry* 7: 212-220. (Frequently attacked by *Scurrula parasitica*.)
- Nandula, V.N., Foy, C.L. and Orcutt, D.M. 1999. Glyphosate for *Orobanche aegyptiaca* control in *Vicia sativa* and *Brassica napus*. *Weed Science* 47: 486-491. (Studying the translocation and fate of glyphosate selectively controlling *O. aegyptiaca* in the naturally glyphosate-tolerant *V. sativa* and genetically engineered glyphosate-resistant *B. napus*.)
- Natilla, A.J. 2000. Ethylene in seed formation and germination. *Seed Science Research* 10: 111-126. (Includes reference to effects of ethylene on *Striga* spp.)
- Norton, D. A. and Ladley, J. J. 1998. Establishment and early growth of *Alepis flavida* in relation to *Nothofagus solandri* branch size. *New Zealand Journal of Botany* 36: 213-217. (Establishment of *Alepis flavida* seedlings was found to be better on smaller diameter branches, about 3mm.)
- Norton, D. A., Ladley, J. J. and Sparrow, A. D. 1997. Development of non-destructive age indices for three New Zealand Loranthaceae mistletoes. *New Zealand Journal of Botany* 35: 337-343. (Describes non-destructive methods of aging *Alepis flavida*, *Ileostylus micranthus* and *Tupeia antarctica*.)
- Otoidobiga, L.C., Vincent, C. and Stewart, R.K. 1998. Relationship between *Smicronyx* spp. population and galling of *Striga hermonthica* (Del.) Benth. *Insect Science and its Application* 18(3): 197-203. (*Smicronyx* adults found on *Striga aspera* before *S. hermonthica* emergence. Each female galled 12-32 seed pods. Damage normally inadequate but might be enhanced with augmentative techniques.)
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